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An efficient protocol has been developed for the synthesis of 2-oxazolines from carboxylic acids and silylated amino alcohols. The advantage of this method was demonstrated by preparing O-silylated amino alcohols. The reaction proceeds via in situ desilvlation of O-silvlated amide followed by cyclization. Studies on silvl deprotection were carried out to explain yield for 2-oxazolines.

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INTRODUCTION

Oxazolines are the most common heterocyclic compounds in many natural products like disorazole and hennoxazole. They are valuable synthetic intermediates 1a, b, as IndPHOX ligands for asymmetric synthesis 1c, d, and also act as protecting groups. For more than a century now, oxazoline rings are the best surrogate to carboxylic acid [2]. 2-Oxazoline, containing natural products, are also known to reduce neurodegenerative diseases [3]. Thus, the oxadiazole moieties are versatile in synthetic utility, that is, synthetic intermediates, ligand scaffolds, chiral auxiliaries, and also chiral catalysts [4,5]. Because of the importance of oxazolines and oxadiazole scaffolds in biological applications like cytotoxic, antitumor, antibacterial, antidepressant, and anti-Alzheimer activities [6,7], many researchers are interested to work on methodologies to simplify the conditions, cost, and time. Some of the molecules such as allosamidin, trehazolin (insecticides), rilmenidine (antihypertensive), A289099 polymerize inhibitor), and bistamide E (Fig. 1) are known to be biologically active due to an oxazoline fragment 7i, i. Ever since oxazolidine-containing natural products have shown promising medicinal value, chiral synthesis of 2oxazolines is the research frontier in discovery as well as medicinal chemistry [8]. A common route to oxazolines is the reaction of an acid chloride with β -amino alcohol; the corresponding hydroxyamide is then treated with thionyl chloride and cyclized with the base via inversion

of the configuration. Several milder approaches have been developed for the cyclization of the β hydroxyamide, including the use of (diethylamino)sulfur trifluoride, Mitsunobu conditions, and PPh₃-CCl₄ [9]. Cyclization of β-hydroxyamide with PPh₃-CCl₄ allows the direct synthesis of oxazolines from carboxylic acids [10]. These methods exhibit less tolerance towards functionalization and causes epimerization [11].

A number of synthetic methods are reported for the synthesis of functionalized oxazoline derivatives from carboxylic acids, aldehydes, nitriles, esters, olefins, carbonyl compounds, and β-hydroxyamides. Pirrung and Tumey described the synthesis of oxazolines using polymer-bound tosyl chloride [12]. The commercially available fluorinating agents DAST and XtalFluor-E were used recently for cyclodehydration of β-hydroxyamides in good yields and without epimerization of α-position [13,14]. Later, Murai et al. reported the synthesis of oxazoline from aldehydes and amino alcohols by using 1,4-Diazabicyclo [2.2.2] octane-DABCO and Chlorosuccinimide-NCS [15]. Chaudhry et al. reported the synthesis of oxazolines from aldehydes and 1,2hydroxyalkyl azides using Lewis acid BF₃-OEt₂ followed by polymer-bonded tosyl hydrazine [16]. Crosignani and Swinnen reported a one-pot protocol using Mukaiyama reagent followed by polymer-bound tosyl chloride [17]. Hazra and his team reported the synthesis of oxazoline derivatives by the reaction of alkenes with NBS in the presence of nitriles and Cu (OTf)₂/Zn (OTf)₂ [18]. Gratia et al. reported the synthesis of oxazolines by the reaction

Figure 1. Examples of 2-oxazoline containing molecules. [Color figure can be viewed at wileyonlinelibrary.com]

of amides and alkenes in the presence of NIS and propionitriles [19].

Orliac and coworkers reported the synthesis of amide by using XtalFluor-E [20]. This group utilized XtalFluor-E as a coupling reagent for optically active substrates without epimerization. Phillips et al. reported the synthesis of oxazolines from β-hydroxyamides using Deoxo-Fluor and DAST [21]. Recently, Brandstatter and coworkers reported the synthesis of 2-oxazolines from silylated protected β-hydroxyamides using XtalFluor-E via in situ desilylation followed by cyclodehydration [22]. But there are hardly any reports towards the synthesis of oxazoline derivatives using O-silylated amino alcohols. As a part of our continuing research interest in methodologies [23], herein, we report the synthesis of substituted 2-oxazoline derivatives using O-silvlated amino alcohols and carboxylic acids using XtalFluor-E under standard conditions (Scheme 1).

RESULTS AND DISCUSSION

The scope of 2-oxazoline synthesis was demonstrated here by taking carboxylic acid 2 and $\beta\text{-hydroxy}$ amino alcohol 1 with tosyl chloride and triethylamine as a base in

tetrahydrofuran solvent. We observed the formation of the desired product (3) in ~10% yield along with side product (4) in 80% yield (Table 1, entry 1). Use of different sulfonyl chlorides like pentafluorosulfonyl chloride and 4nitrobenzene sulfonyl chloride did not alter the yield of the product (Table 1, entries 2 and 3). Then, we switched the coupling reagent by using fluorinating agents like DAST, XtalFluor-E, and Deoxo-Fluor to observe the formation of product 3 in 20 to 25% yield. In these conditions, we observed ester 4 as a side product formed via esterification of intermediate IV and acid in the presence of triethylamine (Table 1, entries 4–6). In order to increase the formation of desired product 3, we carried out the reaction of acid (2) with trimethylsilyl (TMS)-protected and triethylsilyl (TES)-protected β-hydroxyl amino alcohols (1) using XtalFluor-E as reagent and triethylamine as a base at −78°C in tetrahydrofuran solvent (Table 1, entries 7 and 8). Unfortunately, TMS protection was unstable with XtalFluor-E and resulted in ~25% yield, whereas TESprotected β-hydroxyl amino alcohol gave 60% yield towards the product formation 3. Screening of different solvents such as CH₂Cl₂, CHCl₃, 1,4-dioxane, and 1,2-Dichloroethane-DCE resulted in improved yield with dichloromethane facilitating easy workup (Table 1, entry 9). In order to improve the yield of the reaction, various

Scheme 1. One-pot synthesis of 2-oxazolines from carboxylic acids and amino alcohols. [Color figure can be viewed at wileyonlinelibrary.com]

Table 1
Optimization of reaction conditions for the synthesis of oxazoline

S. No.	R	Reagent	Solvent	Temperature	Base	3 (%)
1	Н	Tosyl	THF	0°C	TEA	10
2	Н	Nosyl	THF	0°C	TEA	15
3	Н	Pentafluoro	THF	0°C	TEA	17
4	Н	DAST	THF	-78° C to r.t.	TEA	20
5	Н	XtalFluor-E	THF	78°C to r.t.	TEA	25
6	Н	Deoxo-Fluor	THF	-78° C to r.t.	TEA	20
7	TMS	XtalFluor-E	THF	78°C to r.t.	TEA	25
8	TES	XtalFluor-E	THF	-78° C to r.t.	TEA	60
9	TES	XtalFluor-E	CH ₂ Cl ₂	-78° C to r.t.	TEA	70
10	TES	XtalFluor-E	CHCl ₃	-78° C to r.t.	TEA	65
11	TES	XtalFluor-E	1,4-Dioxane	-78° C to r.t.	TEA	60
12	TES	XtalFluor-E	DCE	-78° C to r.t.	TEA	53
13	TES	XtalFluor-E	CH_2Cl_2	-78° C to r.t.	DIPEA	60
14	TES	XtalFluor-E	CH_2Cl_2	-78° C to r.t.	2,6-Lutidine	65
15	TES	XtalFluor-E	CH_2Cl_2	-78°C to r.t.	Pyridine	80
16	IPDMS	XtalFluor-E	CH_2Cl_2	−78°C to r.t.	Pyridine	85
17	TBS	XtalFluor-E	CH_2Cl_2	78°C to r.t.	Pyridine	50
18	TDS	XtalFluor-E	CH_2Cl_2	78°C to r.t.	Pyridine	40
19	TIPS	XtalFluor-E	CH_2Cl_2	78°C to r.t.	Pyridine	30
20	TBDPS	XtalFluor-E	CH_2Cl_2	78°C to r.t.	Pyridine	30

All reactions were performed with 0.144 mmol of 1 compound, 0.144 mmol of acid 2, 0.432 mmol of base, and 0.312 mmol of XtalFluor-E at -78° C to room temperature.

The best entries during the optimization studies are emphasized in bold.

TES, triethylsilyl; THF, tetrahydrofuran; TMS, trimethylsilyl.

bases like N,N-diisopropyl ethylamine, 2,6-lutidine, and pyridine were screened (Table 1, entries 13, 14, and 15). Among these, pyridine gave a good yield of 80% in CH_2Cl_2 solvent using XtalFluor-E at $-78^{\circ}C$ to ambient temperature. We even observed an increase in yield to 85% on changing the silyl-protecting group from TES to

isopropyldimethylsilyl-IPDMS (Table 1, entry 15 vs 16). Then, we envisioned to study the rate of reaction by changing various silyl-protecting groups such as t-Butyldimethylsilyl-TBS, Thexyldimethylsilyl-TDS, Triisopropylsilyl-TIPS, and t-Butyldiphenylsilyl-TBDPS (Table 1, entries 17–20). Silyl-protecting groups like TBS,

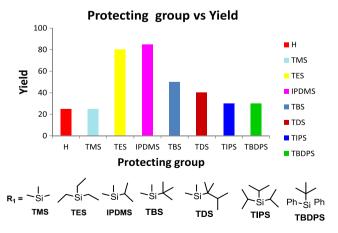


Figure 2. Comparative analysis between % of yield versus protecting group. [Color figure can be viewed at wileyonlinelibrary.com]

TDS, TIPS, and TBDPS decreased the yield for the formation of product 3 due to steric hindrance. Compounds containing bulky protecting groups could not undergo complete deprotection and remain as a major starting material. From the preceding observations, we concluded that the yields were proportional to the rate of silyl deprotection (Fig. 2). The order of silyl group

On the basis of optimized conditions, we synthesized different aryl 2-oxazolines using different acids with IPDMS protected amino alcohols (Scheme 2). Benzoic acid and its derivatives gave excellent yields with optimized conditions of XtalFluor-E (2.2 eq). On the

Scheme 2. Scope of different benzoic acids and different *O*-silylated amino alcohols towards the synthesis of 2-oxazoline derivatives. [Color figure can be viewed at wileyonlinelibrary.com]

Scheme 3. Proposed mechanism. [Color figure can be viewed at wileyonlinelibrary.com]

other hand, the heteroaromatic acids gave moderate to good yields towards the oxazoline formation. The 2-fluoro-5-bromo-substituted amino alcohols on reaction with different acids gave 2-oxazoline derivatives (3a–3g) in moderate to good yields (57–75%). Similarly, simple phenyl amino alcohols on reaction with various acids gave 2-oxazoline derivatives (3h–3m) in good yields. Disubstituted derivatives such as 2-chloro, 4-bromo-substituted amino alcohols also gave good yields towards the formation of oxazoline derivatives (3n–3t).

All synthesized compounds were confirmed by liquid chromatography-mass spectrometry (LC-MS), ¹H NMR, and ¹³C NMR. A possible mechanism for the synthesis of 2-oxazoline derivatives is shown in Scheme 3. XtalFluor-E is attacked by nucleophilic oxygen of aromatic acid 2 to give intermediate I. Nucleophilic attack by nitrogen of amine (i.e., silyl protected amino alcohol) 1 at carbonyl carbon of intermediate I generate intermediate II. Rearrangement followed by elimination of fluoride ion gives intermediate III. Nucleophilic attack of the small fluoride anion leads to a pentavalent silicon centre. The formation of the strong Si–F bond is the driving force for a fast cleavage of O–Si bond. The deprotection of the silyl group *via* fluoride ion from intermediate III gives

intermediate IV, which further reacts with XtalFluor-E followed by cyclization to give compound 3.

CONCLUSION

In summary, we have reported the new protocol for the synthesis of 2-aryl oxazoline derivatives from aromatic acids and O-silylated amino alcohols using XtalFluor-E reagent in excellent yields with a broad range of carboxylic acids, including aromatic, heteroaromatic, aliphatic, and 1°, 2°, and 3° amino alcohols using a slight excess (2.2 equiv.) of the XtalFluor-E. The reaction occurs via in situ formation of β - hydroxy silylated amide followed by the deprotection of the silyl group and then successive cyclization using XtalFluor-E.

EXPERIMENTAL PROCEDURE

2-Bromo-5-[4-(5-bromo-2-fluoro-phenyl)-4-methyl-4,5-dihydro-oxazol-2-yl]-pyridine (3c). To a solution of 2-(5-bromo-2-fluoro-phenyl)-1-((isopropyldimethyl-silyl) oxy) propan-2-amine (0.144 mmol, 1.0 equiv) in dry

dichloromethane (3 mL) was added XtalFluor-E (0.316 mmol, 2.2 equiv) followed by the addition of 6-bromonicotinic acid (0.144 mmol, 1.0 equiv) at $-78^{\circ}\mathrm{C}$. The reaction mixture was slowly warmed to room temperature and stirred for 2 h at room temperature. The reaction mixture was quenched with saturated sodium bicarbonate solution at 0°C and extracted by dichloromethane (3 × 20 mL). The combined organic layers were dried over sodium sulfate and concentrated. The crude product was purified by column chromatography using 20–50% ethyl acetate in hexane as eluent gave pure compounds.

4-(5-Bromo-2-fluoro-phenyl)-4-methyl-2-phenyl-4,5-dihydro-oxazole (3a). White solid. Yield: 36 mg, 75%; 1 H NMR (400 MHz, chloroform-d): δ 8.08–8.05 (m, 2H), 7.98–7.96 (m, 1H), 7.54–7.52 (m, 1H), 7.49–7.45 (m, 2H), 7.39–7.35 (m, 1H), 6.98–6.94 (m, 1H), 4.65 (dd, J = 8.4, 2.8 Hz, 1H), 4.41 (dd, J = 8.8, 2.0 Hz, 1H), 1.67 (s, 3H); 13 C NMR (100 MHz, chloroform-d): δ 163.08, 159.75, 157.32, 136.58, 131.97, 131.51, 130.84, 128.52, 127.61, 127.07, 117.58, 117.35, 117.05, 79.04, 71.14, 28.74; LC-MS: m/z calculated for C_{16} H₁₃BrFNO: 333.02; observed mass 334.2, 336.2 (M + 1, M + 3).

3-[4-(5-Bromo-2-fluoro-phenyl)-4-methyl-4,5-dihydro-oxazol-2-yl]-5-chloro-pyridine (3b). Off white solid. Yield: 33 mg, 62%; 1 H NMR (300 MHz, chloroform-d): δ 8.71–8.70 (m, 1H), 8.13–8.10 (m, 1H), 7.97–7.94 (m, 1H), 7.83–7.79 (m, 1H), 7.39–7.34 (m, 1H), 6.98–6.92 (m, 1H), 4.74 (dd, J = 9.0, 3.6 Hz, 1H), 4.47 (d, J = 8.9 1.9 Hz, 1H), 1.67 (s, 3H); 13 C NMR (100 MHz, chloroform-d): δ 161.87, 161.12, 159.64, 157.21, 142.80, 138.59, 135.96, 131.67, 130.69, 125.88, 123.51, 117.63, 117.10, 79.62, 71.66, 28.79; LC-MS: m/z calculated for $C_{15}H_{11}$ BrClFN₂O: 368.0; observed mass 369.0, 371.0 (M + 1, M + 3).

2-Bromo-5-[4-(5-bromo-2-fluoro-phenyl)-4-methyl-4,5-dihydro-oxazol-2-yl]-pyridine (3c). Off white solid. Yield: 36 mg, 66%; 1 H NMR (400 MHz, DMSO- d_6): δ 8.89 (s, 1H), 8.26–8.23 (m, 1H), 7.82–7.78 (m, 2H), 7.56–7.52 (m, 1H), 7.27–7.22 (m, 1H), 4.73 (dd, J=8.8, 2.8 Hz, 1H), 4.43 (dd, J=8.8, 2.0 Hz, 1H), 1.56 (s, 3H); 13 C NMR (100 MHz, chloroform-d): δ 160.43, 159.65, 157.22, 150.05, 145.37, 138.15, 135.84, 131.68, 130.60, 127.95, 123.04, 117.47, 117.09, 79.29, 71.44, 28.73; LC-MS: m/z calculated for $C_{15}H_{11}Br_2FN_2O$: 411.92; observed mass 413.0, 415.5, 417.0 (M + 1, M + 3, M + 5).

5-[4-(5-Bromo-2-fluoro-phenyl)-4-methyl-4,5-dihydro-oxazol-2-yl]-2-chloro-pyridine (3d). White solid. Yield: 32 mg, 61%; 1 H NMR (400 MHz, DMSO- d_{6}): δ 8.94 (s, 1H), 8.39–8.37 (m, 1H), 7.82–7.80 (m, 1H), 7.69 (d, J = 8.24 Hz, 1H), 7.57–7.54 (m, 1H), 7.29–7.24 (m, 1H), 4.74 (dd, J = 8.8 Hz, 1H), 4.45 (dd, J = 8.8 Hz, 1H), 1.57 (s, 3H); 13 C NMR (75 MHz, DMSO- d_{6}): 162.54, 160.34, 136.89, 132.40, 130.30, 129.17, 127.44, 119.02,

116.81, 78.68, 71.04, 29.62; LC-MS: m/z calculated for $C_{15}H_{11}BrClFN_2O$: 367.97; observed mass 369.0, 371.0 (M + 1, M + 3).

2-[4-(5-Bromo-2-fluoro-phenyl)-4-methyl-4,5-dihydro-oxazol-2-yl]-5-fluoro-pyridine (3e). White solid. Yield: 33 mg, 65%; 1 H NMR (400 MHz, DMSO- 4 6): δ 8.72 (s, 1H), 8.25–8.22 (m, 1H), 7.92–7.87 (m, 1H), 7.81–7.80 (m, 1H), 7.57–7.54 (m, 1H), 7.29–7.24 (m, 1H), 4.74 (dd, 4 J = 8.6 Hz, 1H), 4.44 (dd, 4 J = 8.4 Hz, 1H), 1.57 (s, 3H); LC-MS: 4 Mz calculated for C 4 LT BrF 4 Po: 352.0; observed mass 353.2, 355.2 (M + 1, M + 3).

4-(5-Bromo-2-fluoro-phenyl)-2-(5-methoxypyridin-2-yl)-4-methyl-4,5-dihydrooxazole (3f). White solid. Yield: 34 mg, 65%; 1 H NMR (300 MHz, chloroform-d): δ 8.08 (m, 2H), 8.00 (d, J = 7.8 Hz, 1H), 7.60–7.48 (m, 2H), 7.26–7.21 (m, 1H), 6.86–6.83 (m, 1H), 4.98 (dd, J = 8.6 Hz, 1H), 4.52 (dd, J = 8.58 Hz, 1H), 3.87 (s, 3H), 1.66 (s, 3H); LC-MS: m/z calculated for C₁₆H₁₄BrFN₂O₂: 364.02; observed mass 365.0, 367.0 (M + 1, M + 3).

4-(5-Bromo-2-fluoro-phenyl)-2-(4-fluoro-3-nitrophenyl)-4-methyl-4,5-dihydro-oxazole (3g). White solid. Yield: 32 mg, 57%; 1 H NMR (300 MHz, DMSO- d_6): δ 8.61–8.57 (m, 1H), 8.37–8.33 (m, 1H), 7.82–7.71 (m, 2H), 7.58–7.53 (m, 1H), 7.29–7.23 (m, 1H), 4.77 (dd, J=9.0 2.7 Hz, 1H), 4.47 (dd, J=8.8, 1.5 Hz, 1H), 1.57 (s, 3H); LC-MS: m/z calculated for $C_{16}H_{11}BrF_2N_2O_3$: 396.02; observed mass 397.0, 399.0 (M + 1, M + 3).

2,4-Diphenyl-4,5-dihydrooxazole (3h). White solid. Yield: 30 mg, 80%; 1 H NMR (400 MHz, chloroform-d): δ 8.04 (d, J = 7.2 Hz, 1H), 7.53–7.51 (m, 1H), 7.50–7.43 (m, 2H), 7.38–7.35 (m, 2H), 7.32–7.28 (m, 3H), 5.39 (dd, J = 8.0, 1.2 Hz, 1H), 4.80 (dd, J = 8.4, 0.8 Hz, 1H), 4.28 (t, J = 6.8 Hz, 1H); 13 C NMR (125 MHz, chloroform-d): δ 164.77, 142.41 131.58, 128.79, 128.42, 127.66, 126.79, 74.92, 70.15; LC-MS: m/z calculated for $C_{15}H_{13}NO$: 223.10; observed mass 224.0 (M + 1).

2-(4-Fluoro-phenyl)-4-phenyl-4,5-dihydrooxazole (3i). White solid. Yield: 38 mg, 76%; 1 H NMR (400 MHz, chloroform-d): δ 8.07–8.03 (m, 2H), 7.38–7.31 (m, 2H), 7.30–7.27 (m, 3H), 7.14–7.09 (m, 2H), 5.38 (dd, J = 6.8, 1.2 Hz, 1H), 4.80 (dd, J = 6.8, 1.6 Hz, 1H), 4.28 (t, J = 6.8 Hz, 1H); 13 C NMR (125 MHz, chloroform-d): δ 165.98, 163.89, 163.25, 140.99, 130.64, 128.97, 128.50, 125.86, 124.02, 115.76, 81.37, 63.28; LC-MS: m/z calculated for $C_{15}H_{12}$ FNO: 241.09; observed mass 242.0 (M + 1).

2-(4-Methoxyphenyl)-4-phenyl-4,5-dihydrooxazole (3j). White solid. Yield: 41 mg, 78%; 1 H NMR (400 MHz, chloroform-d): δ 7.99 (dt, J = 8.0, 4.0, 2.0 Hz, 2H), 7.37–7.31 (m, 2H), 7.30–7.26 (m, 3H), 6.96–6.93 (m, 2H), 5.35 (dd, J = 8.0, 0.8 Hz, 1H), 4.77 (dd, J = 8.0, 6.8 Hz, 1H), 4.24 (t, J = 6.8, Hz, 1H), 3.85 (s, 3H); 13 C NMR (125 MHz, chloroform-d): δ 164.57, 162.28,

142.65, 130.25, 128.76, 127.60, 126.80, 120.04, 113.76, 74.83, 70.09, 55.40; LC-MS: m/z calculated for $C_{16}H_{15}NO_2$: 253.11; observed mass 254.0 (M + 1).

2-(4-Nitrophenyl)-4-phenyl-4,5-dihydrooxazole (3k). White solid. Yield: 39 mg, 70%; 1 H NMR (400 MHz, chloroform-d): δ 8.27 (d, J=7.2 Hz, 2H), 8.20 (d, J=6.8 Hz, 2H), 7.39–7.36 (m, 2H), 7.32–7.29 (m, 3H), 5.44 (dd, J=8.0, 7.2 Hz, 1H), 4.86 (dd, J=8.0, 6.8 Hz, 1H), 4.34 (t, J=6.8 Hz, 1H); 13 C NMR (125 MHz, chloroform-d): δ 162.95, 149.60, 141.65, 133.43, 129.57, 128.97, 127.99, 126.78, 123.65, 75.38, 70.47; LC-MS: m/z calculated for $C_{15}H_{12}N_2O_3$: 268.08; observed mass 269.0 (M + 1).

2-(6-Chloropyridin-3-yl)-4-phenyl-4,5-dihydrooxazole (31). White solid. Yield: 39 mg, 72%; 1 H NMR (400 MHz, chloroform-d): δ 8.99 (s, 1H), 8.26 (dd, J = 6.8, 2.0 Hz, 1H), 7.40–7.35 (m, 3H), 7.31–7.27 (m, 3H), 5.39 (dd, J = 8.4 6.8 Hz, 1H), 4.82 (dd, J = 8.4, 6.8 Hz, 1H), 4.30 (t, J = 6.4 Hz, 1H); 13 C NMR (125 MHz, chloroform-d): δ 161.97, 154.33, 149.82 141.68, 138.53, 129.98, 127.99, 126.77, 124.20, 122.82, 75.21, 70.30; LC-MS: m/z calculated for $C_{14}H_{11}ClN_{2}O$: 258.06; observed mass 259.0 (M + 1).

4-Phenyl-2-(trifluoromethyl)-4,5-dihydrooxazole (3m). White solid. Yield: 28 mg, 62%; 1 H NMR (400 MHz, chloroform-d): δ 7.41–7.38 (m, 2H), 7.36–7.32 (m, 1H), 7.26–7.24 (m, 2H), 5.43–5.39 (m, 1H), 4.89 (dd, J = 8.4, 6.8 Hz, 1H), 4.40 (t, J = 6.8 Hz, 1H); LC-MS: m/z calculated for $C_{10}H_8F_3NO$: 215.06; observed mass 217.0 (M + 2). ^{19}F NMR (368 MHz, DMSO- d_6): δ 61.21.

5-(4-Bromo-2-chlorophenyl)-2-(4-fluoro-3-nitrophenyl)-4,5-dihydrooxazole (3o). White solid. Yield: 46 mg, 82%; 1 H NMR (400 MHz, DMSO- d_{6}): δ 8.47 (s, 1H), 8.26–8.22 (m, 2H), 7.68 (m, 1H), 7.54 (d, J = 5.2 Hz, 1H), 7.36 (dd, J = 5.6, 1.6 Hz, 1H), 5.85 (t, J = 8.0 Hz, 1H), 4.50 (t, J = 9.2 Hz, 1H), 3.95 (t, J = 8.4 Hz, 1H); LC-MS: m/z calculated for C₁₅H₉BrClFN₂O₃: 398.00; observed mass 399.0, 401.0 (M + 1, M + 3).

5-(4-Bromo-2-chlorophenyl)-2-(6-bromopyridin-3-yl)-4,5-dihydrooxazole (3p). White solid. Yield 44 mg, 76%; 1 H NMR (400 MHz, DMSO- d_{6}): δ 8.34 (d, J = 2.0 Hz, 1H), 8.28–8.23 (m, 3H), 7.66 (dd, J = 8.0, 2.0 Hz, 1H), 7.38 (dd, J = 6.0, 2.4 Hz, 1H), 5.97 (t, J = 8.0 Hz, 1H), 4.55 (t, J = 8.8 Hz, 1H), 4.06 (dd, J = 9.2, 8.0 Hz, 1H); LC-MS: m/z calculated for $C_{14}H_{9}Br_{2}ClN_{2}O$: 413.88; observed mass 415.0, 417.0, 418.0 (M + 1, M + 3, M + 4).

5-(4-Bromo-2-chlorophenyl)-2-(6-bromopyridin-3-yl)-4,5-dihydrooxazole (3q). White solid. Yield: 42 mg, 70%; 1 H NMR (400 MHz, DMSO- d_{6}): δ 8.59 (d, J = 2.4 Hz, 1H), 8.29 (m, 1H), 8.24 (m, 1H), 8.09 (dd, J = 8.0, 2.8, 1H), 7.63 (d, J = 8.0 Hz, 1H), 7.37 (dd, J = 6.0, 2.4 Hz, 1H), 5.87 (t, J = 8.4, 1H), 4.47 (t, J = 8.8 Hz, 1H), 4.04 (t, J = 8.8 Hz, 1H); LC-MS: m/z calculated for $C_{14}H_{9}Br_{2}ClN_{2}O$: 413.88; observed mass 415.0, 417.0, 418.0 (M + 1, M + 3, M + 4).

5-(4-Bromo-2-chlorophenyl)-2-(5-chloropyridin-3-yl)-4,5-dihydrooxazole (3r). White solid. Yield 38 mg, 72%; 1 H NMR (400 MHz, DMSO- d_{6}): δ 8.27 (s, 1H), 8.22 (d, J = 6.0 Hz, 1H), 7.96–7.94 (m, 2H), 7.69 (d, J = 8.4 Hz, 1H), 7.34–7.32 (m, 1H), 5.90 (t, J = 8.0 Hz, 1H), 4.51 (t, J = 8.8 Hz, 1H), 3.98 (t, J = 8.8 Hz, 1H); LC-MS: m/z calculated for $C_{14}H_{9}BrCl_{2}N_{2}O$: 370.0; observed mass 371.0, 373.0 (M + 1, M + 3).

5-(4-Bromo-2-chlorophenyl)-2-(6-chloropyridin-3-yl)-4,5-dihydrooxazole (3s). White solid. Yield: 38 mg, 72%; 1 H NMR (400 MHz, DMSO- d_{6}): δ 8.27–8.23 (d, 2H), 8.07 (s, 1H), 7.87–7.80 (m, 2H), 7.36 (dd, J=6.0, 2.0 Hz, 1H), 5.89 (t, J=8.4 Hz, 1H), 4.47 (t, J=8.8 Hz, 1H), 3.98 (t, J=8.8 Hz, 1H); LC-MS: m/z calculated for $C_{14}H_{9}BrCl_{2}N_{2}O$: 370.0; observed mass 371.0, 373.0 (M + 1, M + 3).

5-(4-Bromo-2-chlorophenyl)-2-(trifluoromethyl)-4,5-dihydrooxazole (3t). White solid. Yield: 32 mg, 70%; 1 H NMR (400 MHz, DMSO- d_6): δ 7.87–7.80 (m, 2H), 7.54 (t, J = 9.2 Hz, 1H), 5.87 (t, J = 8.8 Hz, 1H), 3.95 (t, J = 8.8 Hz, 1H), 3.47 (t, J = 8.4 Hz, 1H); LC-MS: m/z calculated for $C_{10}H_{6}BrClF_{3}NO$: 326.93; observed mass 328.0, 330.0 (M + 1, M + 3).

2-Benzamido-2-(5-bromo-2-fluoro-phenyl)propyl benzoate (5). White solid. 1 H NMR (400 MHz, CDCl₃): δ 8.03 (dd, J = 1.2, 8.4 Hz, 2H), 7.80 (dd, J =, 8.4,1.2 Hz, 2H), 7.63–7.60 (m, 2H), 7.59–7.57 (m, 6H), 6.94 (dd, J = 8.8, 12.0 Hz, 1H), 4.97 (d, J = 11.2 Hz, 1H), 4.63 (d, J = 11.2 Hz, 1H), 1.58 (s, 3H); LC-MS: m/z calculated for C₂₃H₁₉BrFNO₃: 455.05; observed mass 456.2, 458.2 (M + 1, M + 3).

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REFERENCES AND NOTES

[1] (a) Wipf, P.; Venkatraman, S. J. Org Chem 1995, 60, 7224; (b) Ousmer, M.; Braun, N. A.; Ciufolini, M. A. Org Lett 2001, 5, 765; (c) McManus, H. A.; Guiry, P. J Chem Rev 2004, 104, 4151; (d) Wang, Y.; Hamalainen, A.; Tois, J.; Franzen, R. Tetrahedron Asymmetry 2010, 21, 2376; (e) Wang, Y.; Hamalainen, A.; Tois, J. E.; Franzen, R. Tetrahedron Asymmetry 2011, 22, 524; (f) Greene, T. W.; Wuts, P. G. M. Protecting Groups in Organic Synthesis, 2nd ed.; John Wiley & Sons: New York, 1991.

[2] (a) Wang, Y.; Sauer, D. R.; Djuric, S. W. Tetrahedron Lett 2006, 47, 105; (b) Gant, T. G.; Meyers, A. I. Tetrahedron 1994, 50, 2297.

[3] (a) Wipf, P.; Venkatraman, S. Synlett 1997 1; (b) Rodriguez, A. D.; Ramirez, C.; Rodriguez, I. I.; Gonzalez, E. Org Lett 1999, 1, 527; (c) Campiani, G.; Angelis, M. D.; Armaroli, S.; Fattorusso, C.; Catalanotti, B.; Ramunno, A.; Nacci, V.; Novellino, E.; Grewer, C.;

- Ionescu, D.; Rauen, T.; Griffiths, R.; Sinclair, C.; Fumagalli, E.; Mennini, T. J Med Chem 2001, 44, 2507.
- [4] (a) Greene, T. W.; Wuts, P. G. M. Protective Groups in Organic Synthesis, 2nd ed.; John Wiley & Sons, 1991 265–266 and 433–436; (b) Corey, E. J.; Ishihara, K. Tetrahedron Lett 1992, 33, 6807; (c) Meyers, A. I.; Miheich, E. D. Angew Chem Int Ed 1976, 15, 270.
- [5] (a) Meyers, A. I. J Heterocyclic Chem 1998, 35, 991; (b) Go'mez, M.; Muller, G.; Rocamora, M. Coord Chem Rev 1999, 193, 769; (c) Wipf, P.; Wang, X. Org Lett 2002, 4, 1197; (d) Johns, B. A. PCT Int Appl. WO 2004101512; (e) Piatnitski, E.; Kiselyov, A.; Doody, J.; Hadari, Y.; Ouyang, S.; Chen, X. PCT Int Appl. WO 2004052280
- [6] (a) Adelstein, G. W.; Yen, C. H.; Dajani, E. Z.; Bianchi, R. G. J Med Chem 1976, 19, 1221; (b) Dogan, H. N.; Duran, A.; Rollas, S.; Sener, G.; Uysal, M. K.; Gulen, D. Bioorg Med Chem 2002, 10, 2893; (c) Palaska, E.; Sahin, G.; Kelicen, P.; Durlu, N.; Tugba; Altinok, G. IL Farmaco 2002, 57, 101; (d) Brown, P.; Best, D. J.; Broom, N. J. P.; Cassels, R.; O'Hanlon, P. J.; Mitchell, T. J.; Osborne, N. F.; Wilson, J. M. J Med Chem 1997, 40, 2563; (e) Savitha, B.; Reddy, E. K.; Parthasarathi, D.; Rajeesh, P.; Sajith, A. M.; Ananda kumar, C. S.; Haridas, K. R.; Ali Padushaa, M. S. J Heterocyclic Chem 2018, 55, 2277.
- [7] (a) Jansen, R.; Irschik, H.; Reichenbach, H.; Wray, V.; Hoefle, G. Liebigs Ann Chem 1994, 1994, 759; (b) Celanire, S.; Talaga, P.; Leurs, R.; Denonne, F.; Timmerman, H.; Lebon, F. Patent no. WO 2006103057 A1, 2006; (c) Onishi, H. R.; Pelak, B. A.; Silver, L. L.; Kahan, F. M.; Chen, M.-H.; Patchett, A. A.; Galloway, S. M.; Hyland, S. A.; Anderson, M. S.; Raetz, C. R. H. Science 1996, 274, 980; (d) Tsukamoto, M.; Murooka, K.; Nakajima, S.; Abe, S.; Suzuki, H.; Hirano, K.; Kondo, H.; Kojiri, K.; Suda, H. J Antibiot 1997, 50, 815; (e) Vizi, E. S. Med Res Rev 1986, 6, 431; (f) Gross, J. L.; Robichaud, A. J.; Mazzacani, A.; Williams, M. J. Patent no. US 0023707, 2009; (g) Bergeron, R. J.; Xin, M. G.; Weimar, W. R.; Smith, R. E.; Wiegand, J. J Med Chem 2001, 44, 2469; (h) Hopkins, C. D.; Wipf, P. Nat Prod Rep 2009, 26, 585; (i) Tsuda, M.; Yamakawa, M.; Oka, S.; Tanaka, Y.; Hoshino, Y.; Mikami, Y.; Sato, A.; Fujiwara, H.; Ohizumi, Y.; Kobayashi, J. J Nat Prod 2005, 68, 462.
- [8] (a) Parsons, R. L. Jr.; Heathcock, C. H. J Org Chem 1994, 59, 4733; (b) Wipf, P. In Alkaloids: Chemical and Biological PerspectivesPelletier, S. W. Ed.; Pergamon: New York, 1998, p 187; (c) Perez, L. J.; Faulkner, D. J J Nat Prod 2003, 66, 247; (d) Kim, M. Y.; Vankayalapati, H.; Shin-ya, K.; Wierzba, K.; Hurley, L. H. J J Am Chem Soc 2002, 124, 2098; (e) Sajith, A. M.; Abdul Khader, K. K.; Muralidharan, A.; Ali, Padusha, M. S.; Nagaswarupa, H. P. J Heterocyclic Chem 2015, 52, 1748.
- [9] (a) Meyers, A. I. J Org Chem 2005, 70, 6137; (b) Reuman, M.; Meyers, A. I. Tetrahedron 1985, 41, 837.

- [10] (a) Vorbruggen, H.; Krolikiewicz, K. Tetrahedron 1993, 49, 9353; (b) Rajaram, S.; Sigman, M. S. Org Lett 2002, 4, 3399.
- [11] (a) Wipf, P.; Miller, C. P. Tetrahedron Lett 1992, 33, 6267; (b) Metcalf, T. A.; Simionescu, R.; Hudlicky, T. J Org Chem 2010, 75, 3447.
 - [12] Pirrung, M. C.; Tumey, L. N. J Comb Chem 2000, 2, 675.
- [13] Burrell, G.; Evans, J. M.; Jones, G. E.; Stemp, G. Tetrahedron Lett 1990, 31, 3649.
- [14] L'Heureux, A.; Beaulieu, F.; Bennett, C.; Bill, D. R.; Clayton, S.; LaFlamme, F.; Mirmehrabi, M.; Tadayon, S.; Tovell, D.; Couturier, M. J Org Chem 2010, 75, 3401.
- [15] Murai, K.; Takahara, Y.; Matsushita, T.; Komatsu, H.; Fujioka, H. Org Lett 2010, 12, 15, 3456.
- [16] Chaudhry, P.; Schoenen, F.; Neuenswander, B.; Lushington, G. H.; Aube, J. J Comb Chem 2007, 9, 473.
 - [17] Crosignani, S.; Swinnen, D. J Comb Chem 2005, 7, 688.
- [18] Hajra, S.; Bar, S.; Sinha, D.; Maji, B. J Org Chem 2008, 73, 4320.
- [19] (a) Gratia, S. S.; Vigneau, E. S.; Eltayeb, S.; Patel, K.; Meyerhoefer, T. J.; Kershaw, S.; Huang, V.; Castro, M. D. Tetrahedron Lett 2014, 55, 448; (b) Cyrous, O.; Kangani; Kelley D E Tetrahedron Lett 2005, 46, 8917.
- [20] Orliac, A.; Pardo, D. G.; Bombrun, A.; Cossy, J. Org Lett 2013, 15, 4, 902.
- [21] Phillips, A. J.; Uto, Y.; Wipf, P.; Reno, M. J.; Williams, D. Org Lett 2000, 2, 1165.
- [22] Brandstatter, M.; Roth, F.; Luedtke, N. W. J Org Chem 2015, 80, 40.
- [23] (a) Reddy, E. K.; Chandran, R.; Sajith, A. M.; Dileep, K. V.; Sadasivan, C.; Anwar, S. RSC Adv 2016, 6, 77431; (b) Reddy, E. K.; Chandran, R.; Mantosh, K.; Sajith, A. M.; Omkumar, R. V.; Sadasivan, C.; Anwar, S. Eur J Med Chem 2017, 139, 367; (c) Babu, G. V.; Settipalli, P. C.; Reddy, E. K.; Anwar, S. Eur J Org Chem 2019, 2019, 2234; (d) Prasad, T. N.; Reddy, E. K.; Babu, G. V.; Basha, S. F.; Anwar, S. Synth Commun 2019, 49, 1277.

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